

## Intracellular Screen To Identify Metagenomic Clones That Induce or Inhibit a Quorum-Sensing Biosensor

Lynn L. Williamson,<sup>1,2</sup> Bradley R. Borlee,<sup>2</sup> Patrick D. Schloss,<sup>2</sup> Changhui Guan,<sup>2</sup>  
Heather K. Allen,<sup>2,3</sup> and Jo Handelsman<sup>2\*</sup>

Department of Bacteriology,<sup>1</sup> Department of Plant Pathology,<sup>2</sup> and Microbiology Doctoral Training Program,<sup>3</sup> University of Wisconsin—Madison, Madison, Wisconsin 53706

Received 7 February 2005/Accepted 18 May 2005

**The goal of this study was to design and evaluate a rapid screen to identify metagenomic clones that produce biologically active small molecules. We built metagenomic libraries with DNA from soil on the floodplain of the Tanana River in Alaska. We extracted DNA directly from the soil and cloned it into fosmid and bacterial artificial chromosome vectors, constructing eight metagenomic libraries that contain 53,000 clones with inserts ranging from 1 to 190 kb. To identify clones of interest, we designed a high throughput “intracellular” screen, designated METREX, in which metagenomic DNA is in a host cell containing a biosensor for compounds that induce bacterial quorum sensing. If the metagenomic clone produces a quorum-sensing inducer, the cell produces green fluorescent protein (GFP) and can be identified by fluorescence microscopy or captured by fluorescence-activated cell sorting. Our initial screen identified 11 clones that induce and two that inhibit expression of GFP. The intracellular screen detected quorum-sensing inducers among metagenomic clones that a traditional overlay screen would not. One inducing clone carries a LuxI homologue that directs the synthesis of an *N*-acyl homoserine lactone quorum-sensing signal molecule. The LuxI homologue has 62% amino acid sequence identity to its closest match in GenBank, AmfI from *Pseudomonas fluorescens*, and is on a 78-kb insert that contains 67 open reading frames. Another inducing clone carries a gene with homology to homocitrate synthase. Our results demonstrate the power of an intracellular screen to identify functionally active clones and biologically active small molecules in metagenomic libraries.**

Soil is densely populated with microorganisms that produce many small bioactive molecules, including antibiotics, anticancer compounds, immunosuppressive agents, insecticides, and other molecules that are important in nature and useful to humans (22, 45, 46, 48). Most of the known molecules of these classes are derived from the cultured microorganisms in soil, which represent <1% of the soil community (63). Soil is likely to contain >6,000 species per gram (7) and therefore probably contains more extensive functional diversity than is represented by cultured microorganisms (6, 24, 25).

Metagenomics, or the analysis of the collective genomes of an assemblage of organisms (22, 52), provides a strategy to find bioactive compounds from among uncultured organisms. Metagenomic libraries are constructed from DNA extracted directly from an environmental sample. Functional profiling of metagenomic libraries, in which all clones in a library that express a certain function are identified and then sequenced, may provide insight into the genomic context of the genes encoding the function and the phylogeny of the organisms that contain these genes (24, 50, 54). This approach has recovered novel biocatalysts and bioactive compounds, and has expanded our knowledge of uncultured microorganisms (for reviews, see references 8, 25, and 50). However, the outstanding challenges in functional metagenomics include screening the massive libraries needed to represent all of the genomes in an environmental sample and overcoming barriers to heterologous gene

expression in the host species harboring the library. These challenges are significant barriers in identifying small molecules such as antibiotics and signal compounds (24, 26, 38, 60, 65). Typically, these have been identified in screens involving overlays with indicator organisms, but these screens are not sufficiently sensitive, only detect exported molecules, and are too laborious to apply easily to libraries containing millions of clones.

Our focus is on discovering small molecules in members of soil communities that have not been well studied. The site of interest in the present study is a nonpermafrost soil supporting a balsam poplar stand on a floodplain island in the Tanana River in central Alaska. The average daily soil temperature remains below 12°C (42). Unlike microorganisms in many soils of temperate climates, microorganisms in the extreme terrestrial environments of Alaska have not been well studied. This extreme environment might be home to microorganisms that produce novel biologically active molecules, as indicated by the novelty of organisms in other cold environments. For example, a study of a Siberian tundra soil using culture-independent methods revealed 16S rRNA sequences that diverge deeply from known sequences and may represent previously undescribed divisions of bacteria (69). Moreover, substantial metabolic activity has been measured in permafrost sediments (51), an arctic glacier (56), and mountain snow (55).

To develop an intracellular screen for small molecules, we sought a system that responds to diverse molecules and is found in diverse organisms and environments. Quorum sensing satisfied these requirements. It is a process of cell-cell communication mediated by small signal molecules that enables bac-

\* Corresponding author. Mailing address: Department of Plant Pathology, University of Wisconsin—Madison, 1630 Linden Dr., Madison, WI 53706. Phone: (608) 263-8783. Fax: (608) 265-5289. E-mail: joh@plantpath.wisc.edu.

TABLE 1. Bacterial strains and plasmids

Strain or plasmid	Relevant characteristic(s) <sup>a</sup>	Source or reference
Biosensor strains and plasmids		
<i>Agrobacterium tumefaciens</i> KYC55(pJZ410, pJZ384, pJZ372)	KYC55: R10 derivative lacking pTiR10. pJZ410, pJZ384, pJZ372: T7 expression system to express TraR	5, 70
<i>Agrobacterium tumefaciens</i> NT1 (pTiC58Δ <i>accR</i> )	Acyl-HSL producer	3
<i>Chromobacterium violaceum</i> CV026	Acyl-HSL-negative mutant of ATCC 31532; most sensitive to <i>N</i> -hexanoyl-L-HSL	41
<i>Escherichia coli</i> MT102 (pJBA132), referred to as reporter strain "JB525"	MT102: <i>araD139 (ara-leu)</i> 7697 Δ <i>lac thi hsdR</i> . pJBA132: pME6031 carrying <i>luxR</i> -P <sub>luxI</sub> -RBSII- <i>gfp</i> (ASV)-T <sub>0</sub> -T <sub>1</sub> ; Tet <sup>r</sup> ; based on components of the <i>V. fischeri lux</i> quorum-sensing system, it is most sensitive to <i>N</i> -(3-oxohexanoyl)-L-HSL	1
<i>Pseudomonas aeruginosa</i> PAO1	Acyl-HSL producer	<i>Pseudomonas</i> Genetic Stock Center
<i>Pseudomonas putida</i> F117 (pKR-C12)	F117: acyl-HSL-negative derivative of IsoF, PpuI <sup>-</sup> . pKR-C12: pBBR1MCS-5 carrying P <sub>lasB</sub> - <i>gfp</i> (ASV)-P <sub>lac</sub> - <i>lasR</i> ; Gm <sup>r</sup> ; based on components of the <i>P. aeruginosa las</i> quorum-sensing system, it is most sensitive to <i>N</i> -(3-oxododecanoyl)-L-HSL and other long-chain acyl-HSL molecules	57
<i>Pseudomonas putida</i> IsoF	Acyl-HSL producer	57
<i>E. coli</i> hosts for library construction or cloning		
DH5α	<i>recA1</i> and <i>endA1</i> cloning strain	Lab stock
TransforMAX 300 EPI300	<i>trfA</i> <sup>+</sup> host for inducible copy number	Epicentre
Vectors		
pSuperBAC	High-copy-inducible BAC cloning vector; Cm <sup>r</sup>	23
pCC1BAC	High-copy-inducible BAC cloning vector; Cm <sup>r</sup>	Epicentre
pCC1FOS	High-copy-inducible fosmid cloning vector; Cm <sup>r</sup>	Epicentre
pGEM-T	Cloning vector	Promega
pGEM-3Zf(+)	Cloning vector	Promega

<sup>a</sup> Tet<sup>r</sup>, tetracycline resistance; Gm<sup>r</sup>, gentamicin resistance; Cm<sup>r</sup>, chloramphenicol resistance.

teria to sense their own cell density. Quorum sensing has emerged as a central aspect of microbial life, and therefore diverse, as-yet-uncultured organisms are likely to produce compounds that regulate quorum sensing. Structural variation in quorum-sensing inducers provides specificity for regulation of many ecologically important functions, which include luminescence (12, 27), virulence (30, 32, 47, 49, 66), and antibiotic production (2, 43, 67). Molecules that have been shown to inhibit quorum sensing include furanones (18), lactonases (10), acylases (33), and long-chain acyl-homoserine lactones (HSLs) (41).

Quorum sensing was discovered in *Vibrio fischeri*, in which LuxI directs the synthesis of the acyl-HSL signal molecule (12, 44). When the signal molecule reaches sufficient concentration in the cell with the transcriptional activator, LuxR, it activates transcription of target genes, including *luxI*, in an amplification loop (15). Diverse small molecules other than acyl-HSL molecules activate or antagonize *lux*-based biosensors (9, 16, 20, 28, 40). Of particular interest is the finding that sublethal concentrations of antibiotics such as rifampin and erythromycin induce quorum-sensing-regulated gene expression (20). Based on these features, we chose the *luxI-luxR* system as the basis for an intracellular screen.

Here we describe construction and screening of metagenomic libraries from Alaskan soil microorganisms. We present and evaluate an intracellular screen, designated METREX, for metagenomic libraries in which the biosensor that detects active clones is inside the same cell as the metagenomic DNA.

## MATERIALS AND METHODS

**Soil.** Soil was collected from islands, BP1 and BP3, in the National Science Foundation Long-Term Ecological Research site at Bonanza Creek Experimental Forest near Fairbanks, Alaska, in October 2001, May 2002, and October 2002. Organic and mineral layers from soil cores were treated as separate samples. The textures, organic matter contents, and pHs of representative soil cores were determined by standard methods at the Soil and Plant Analysis Lab, University of Wisconsin—Madison (see Table 2). Soil was put through a 5-mm sieve to remove roots and large particulates and stored at 4°C until use.

**Bacterial strains and culture conditions.** The bacterial strains and plasmids used in the present study are listed in Table 1. Reporter strain JB525 (*Escherichia coli* MT102 containing recombinant plasmid pJBA132) obtained from M. Givskov, Technical University of Denmark, was used for the intracellular screen (1). Electrocompetent *E. coli* TransforMax EPI300 (Epicentre, Madison, WI) was used for library construction, and *E. coli* DH5α(pGEM-3Zf(+)) and pGEM-T (Promega, Madison, WI) was used for subcloning. All *E. coli* strains were grown in Luria-Bertani (LB) medium at 37°C, except for reporter strain JB525, which was grown in LB amended with 5 g of NaCl/liter. *Agrobacterium tumefaciens* acyl-HSL indicator strains KYC55(pJZ410)(pJZ384)(pJZ372) (70) and NT1(pTiC58Δ*accR*) (3) were grown in a yeast extract-mannitol (AMA) (34) broth medium at 28°C. *Pseudomonas putida* F117(pKR-C12) (57), *Pseudomonas aeruginosa* PAO1, and *Chromobacterium violaceum* CV026 (41) acyl-HSL indicator strains were maintained on LB agar at 28°C. Where appropriate, media were amended with chloramphenicol (Cm) at a concentration of 10 to 20 μg ml<sup>-1</sup>, tetracycline (Tet) at a concentration of 20 μg ml<sup>-1</sup>, or ampicillin at a concentration of 150 μg ml<sup>-1</sup>.

**DNA extraction and preparation.** Two procedures were used to extract DNA: direct lysis within the soil matrix (direct extraction) and lysis of cells that were separated from the soil matrix by differential centrifugation prior to lysis (cell separation). The direct extraction procedure, used for libraries AK1 through AK4, was based on the method of Rondon et al. with modifications as follows (52). The cell separation procedure, used for libraries AK5 through AK8, was performed as previously described (37, 64). For the direct extraction procedure used for libraries AK1 through AK4, 100 g of soil was mixed with 150 ml of extraction buffer (68) in centrifuge tubes and subjected to two freeze-thaw cycles.

TABLE 2. Libraries constructed with DNA from Alaskan soil

Library	Soil properties			Vector	No. of clones	Avg insert size (kb)	No. of bases (Mb) <sup>d</sup>
	Layer	Texture <sup>a</sup> (% sand/silt/clay)	%OM <sup>b</sup>				
AK1	Organic	15/80/05	11	6.7	pSuperBAC	22,200	222
AK2	Mineral	08/84/08	8	7.5	pSuperBAC	4,600	23
AK3	Mineral	19/77/04	3	6.8	pCC1BAC	2,500	33
AK4	Organic	16/74/10	40	5.5	pSuperBAC	16,800	185
AK5	Organic	16/74/10	40	5.5	pSuperBAC	1,400	46
AK6	Organic	16/74/10	40	5.5	pCC1FOS	300	12
AK7	Organic	16/74/10	40	5.5	pCC1BAC	2,300	108
AK8	Organic	16/74/10	40	5.5	pCC1BAC	2,700	24
Total						52,800	653

<sup>a</sup> Texture is the percent sand/silt/clay determined by physical analysis of soil prepared with dispersing solution and measured with a hydrometer.

<sup>b</sup> %OM is the percent organic matter determined by loss of weight on ignition at 360°C for 2 h.

<sup>c</sup> Prepared as a 1:1.3 (vol/vol) soil-to-water mixture and measured with a glass electrode.

<sup>d</sup> Total amount of DNA cloned in the library, calculated as the product of the average insert size and the number of clones in the library.

Each cycle consisted of immersion for 30 min in a slush of crushed dry ice and isopropanol, followed by immersion of the sample in a 65°C water bath for 30 min. Then, 18 ml of 20% (wt/vol) sodium dodecyl sulfate plus 9 ml of 5 M guanidine isothiocyanate was added to the warmed soil solution, which was subsequently incubated at 65°C for 2 h with occasional gentle mixing. Supernatants, collected after centrifuging the samples at 15,000 × g for 20 min at 10°C, were combined with 25 ml of chloroform-isoamyl alcohol (24:1 [vol/vol]) and mixed gently for 10 min. The aqueous phase was recovered by centrifugation at 15,000 × g for 20 min at 10°C and precipitated with a 0.7 volume of isopropanol at room temperature for 20 min. The DNA was pelleted by centrifugation at 15,000 × g for 40 min at 10°C, resuspended in 2 ml of 10 mM Tris (pH 8.0) plus 10 mM EDTA (pH 8.0), and extracted by adding an equal volume of Tris-buffered phenol-chloroform (pH 8.0) and centrifuging the mixture at ≥10,000 × g (14,000 rpm) in an Eppendorf 5417C tabletop centrifuge for 10 min at room temperature. The aqueous layer containing the DNA was removed and extracted again with chloroform-isoamyl alcohol (24:1 [vol/vol]). The DNA suspension was stored at -80°C until it was purified by electrophoresis in a low-melting-temperature agarose gel (LMP Preparative Grade for Large Fragments [Promega] or GenePureLowMelt GQA [ISC BioExpress, Kaysville, UT]). Sections containing DNA were cut from the unstained gel and treated with GELase (Epicenter) or AgarACE (Promega) according to the manufacturers' directions.

**Library construction.** We constructed four bacterial artificial chromosome (BAC) libraries (AK1, -2, -4, and -5) in the vector pSuperBAC (23), three BAC libraries (AK3, -7, and -8) in CopyControl pCC1BAC (Epicenter), and one fosmid library (AK6) in CopyControl pCC1FOS (Epicenter) (Table 2). The pSuperBAC was prepared according to the method of Gillespie et al. (17) with minor modifications as follows. pSuperBAC was isolated and purified with QIAprep Miniprep (QIAGEN, Valencia, CA) and digested with 10 U of HindIII (Promega). The DNA was dephosphorylated by using shrimp alkaline phosphatase (Promega), which was then heat inactivated according to the manufacturer's conditions. The dephosphorylated product was precipitated, self-ligated, and gel purified as previously described (17) and extracted by using the QIAquick Gel Extraction Kit (QIAGEN).

For libraries AK1 through AK4, the purified soil DNA was partially digested with ca. 25 U of HindIII per 1 μg of DNA at 37°C for 1 h. For AK5, AK7, and AK8, agarose plugs containing DNA, resulting from the cell separation extraction method, were soaked in TE (10 mM Tris, 1 mM EDTA [pH 8.0]) for 1 h on ice and then in buffer solution (Sau3AI buffer and acetylated bovine serum albumin [Promega] plus polyamines as described previously [http://www.tree.caltech.edu/protocols/BAC\_lib\_construction.html]) for 30 min on ice, using about 200 μl of buffer solution for a 400-μg plug. To achieve partial digests, ca. 50 U of Sau3AI was added, and the plugs were incubated for 30 min on ice and then 2 h at 37°C. The reaction was stopped by adding a 0.1 volume of 0.5 M EDTA and placing the mixture on ice for 20 min. The plugs were soaked in TE prior to size fractionation. Partially digested DNA was loaded onto a 1% low-melting-point agarose pulsed-field gel (Bio-Rad CHEFMapper; Bio-Rad Laboratories, Hercules, CA) at 6 V/cm with a 0.5- to 15-s switch time, a linear ramping factor, and a 120° included angle for 10 h in 1× TAE buffer (50× stock solution is 2.5 M Tris, 50 mM EDTA, adjusted to pH 7.5 to 7.8 with glacial acetic acid).

DNA larger than 50 kb was excised and extracted from the gel with GELase or AgarACE as described above.

DNA ligation reactions contained an insert/vector ratio of 1:10 for pSuperBAC or 1:1 for CopyControl pCC1BAC. Vector and insert were heated to 55°C for 10 min and then cooled at room temperature 15 min prior to adding 3 U of T4 DNA ligase and buffer (Promega). Ligation reactions were incubated overnight at 16°C. Transformation was performed as described previously (17) and plated on LB medium containing Cm at 10 μg ml<sup>-1</sup>. Libraries were evaluated by restriction digest analysis of randomly selected clones using NotI. Colonies were pooled by scraping them from plates into LB containing Cm and mixing them with an equal volume of 14% dimethyl sulfoxide, followed by storage at -80°C.

To construct AK6, purified DNA (2.5 μg) was enzymatically treated to prepare 5' phosphorylated blunt ends and ligated by using a CopyControl Fosmid Library Production Kit (Epicenter). After packaging into lambda phages, infected cells were plated and stored as described above for the BAC libraries.

**Intracellular screen.** In this screen, plasmid DNA from pooled metagenomic clones (Table 2) was transferred into cells containing the biosensor plasmid pJBA132 (provided by M. Givskov) (Fig. 1), and clones were detected by fluorescence-activated cell sorting (FACS) or fluorescence microscopy. A pool of all clones from libraries AK1 through AK8 was prepared and then incubated in SOB (0.5% yeast extract, 2.0% tryptone, 0.05% NaCl, 10 mM MgCl<sub>2</sub>, 2.5 mM KCl) for 2 h at 37°C, and the plasmid DNA was isolated by using QIAprep Miniprep (QIAGEN) and precipitated with isopropanol (53). From this preparation, 50 ng of DNA was transformed into reporter strain JB525 (*E. coli* MT102 containing pJBA132) using the conditions provided in the description of library construction. After 1 h of recovery in SOB at 37°C with shaking, 1:300 dilutions were prepared by using 10 μl of recovery mix in 3-ml total volume of SOB containing Cm and Tet and 50 and 80 nM, respectively, *N*-(3-oxohexanoyl)-L-HSL. The same host carrying the empty vector served as a negative control. The dilutions were incubated at 28°C for 5 h and sorted into green fluorescent protein (GFP)-expressing and non-GFP-expressing fractions by using an EPICS Elite flow cytometer/cell sorter (Coulter Corp., Fullerton, CA) as previously described (11). Non-GFP-expressing cells were collected into 1 ml of SOB containing Cm and Tet and cultured on plates containing the same medium.

Alternatively, plasmid DNA isolated from pooled metagenomic clones (Table 2) was transformed into reporter strain JB525 and plated at a density sufficiently low to facilitate visualizing single colonies on LB medium containing Cm and Tet. After overnight incubation at 37°C, colonies were screened for fluorescence by using a fluorescence microscope (Leica MZ FLIII; Leica Microsystems, Deerfield, IL) with a GFP filter (excitation wavelength, 470/40 nm; barrier filter, 525/50 nm). The reporter strain containing empty vector provided a negative control. This screen was repeated three times.

All clones that were positive in the initial screens were retested in a fresh host background. The plasmids (BAC/fosmid plasmid and biosensor plasmid) were isolated, retransformed into *E. coli* TransformMax EPI300, and plated on LB medium containing Cm to select for retransformants containing the BAC or fosmid plasmid. Resulting colonies were tested on LB medium containing Tet to verify and eliminate colonies that contained both the biosensor and BAC/fosmid plasmids. Isolated BAC/fosmid plasmids were retransformed into a fresh re-

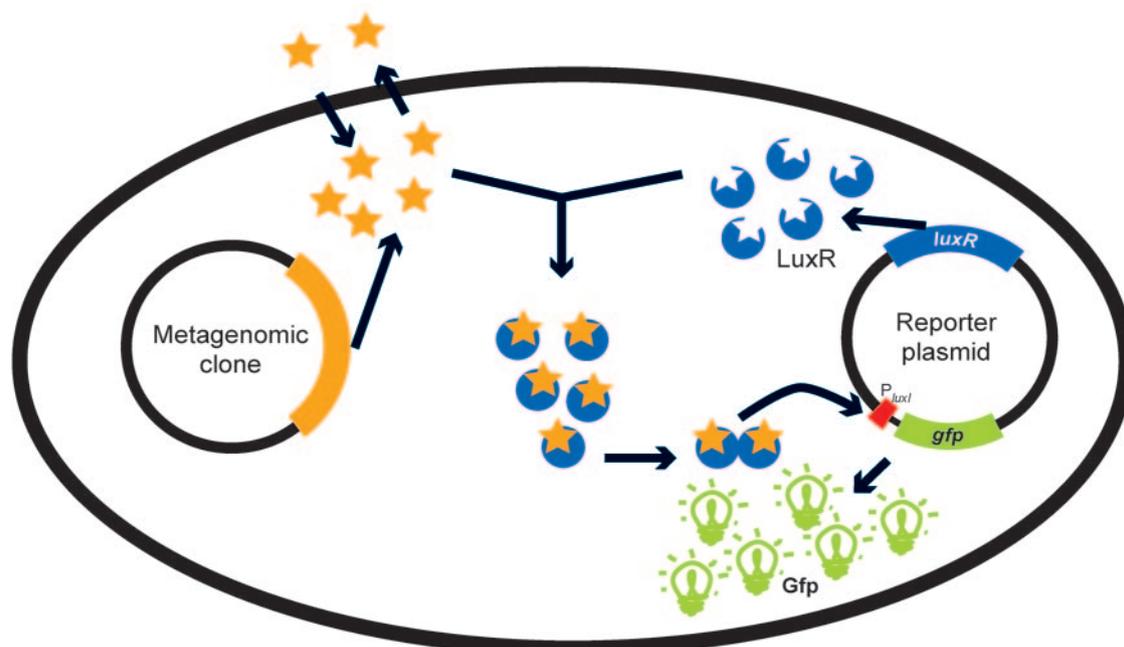


FIG. 1. In this intracellular screen, the biosensor that detects active clones is inside the same cell as the metagenomic DNA. The biosensor detects small diffusible signal molecules that induce quorum sensing. When the signal molecules reach a sufficient concentration, they bind the LuxR transcriptional activator which activates  $P_{luxI}$  and induces expression of target genes, in this case GFP. The biosensor plasmid was received from Michael Givskov, Technical University, Lyngby, Denmark.

porter strain JB525, and the phenotype of the transformants was assessed to determine whether the cloned DNA was responsible for the phenotype. Plasmids isolated from the original colonies and from the retransformed colonies were digested with NotI, and the restriction digest patterns were compared by pulsed-field gel electrophoresis.

**Specificity of biosensor.** Genomic DNA was isolated with the Easy-DNA kit (Invitrogen, Carlsbad, CA) from *P. aeruginosa* PAO1, *P. putida* IsoF, and *A. tumefaciens* NT1. Primers were designed to amplify *lasI*, *ppuI*, and *traI* based on published sequences (the accession numbers are AE004572 [59], AY115588 [58], and L22207 [29], respectively) (*lasI* forward, 5'-ATC GTA CAA ATT GGT CGG-3'; *lasI* reverse, 5'-TGA AAC CGC CAG TCG CTG-3'; *ppuI* forward, 5'-CTT ATC TCC ATT GAT CGC C-3'; *ppuI* reverse, 5'-ATG GCA GTT ATT CTT GAT-3'; *traI* forward, 5'-CGG ATA CTC ACA GTT TCG-3'; *traI* reverse, 5'-CGC GGC ACT CCT GAT CGC-3'). PCR amplification of acyl-HSL synthase genes (50- $\mu$ l reaction volume) used 100 ng of genomic DNA, a 200  $\mu$ M concentration of each deoxynucleoside triphosphate, 2.5 U of *Taq* polymerase (Promega), and 20 pmol of each forward and reverse primer. Reactions were performed in an Eppendorf Mastercycler gradient thermocycler (Eppendorf, Hamburg, Germany) using 1 min of denaturation at 95°C and then 40 cycles of 30 s at 95°C, 90 s at 55°C, and 150 s at 72°C, followed by 5 min of extension at 72°C.

The resulting products were gel purified with the QIAEX II gel extraction kit (QIAGEN) and cloned into the TA cloning vector pGEM-T (Promega). Ligation products were transformed into electrocompetent JB525 and, after overnight incubation at 37°C, colonies were screened for fluorescence using a fluorescence microscope to confirm activation of the biosensor.

**Overlay assay to detect bioactive compounds produced by metagenomic clones.** To compare the sensitivity of the intracellular screen to an overlay screen, active clones detected by the intracellular screen were plated at a density sufficiently low to visualize single colonies on LB medium, incubated overnight at 37°C, and overlaid with 6 ml of soft agar (LB with 0.005 g of agarose ml<sup>-1</sup>) that had been inoculated with 0.05 ml of an overnight culture of reporter strain JB525. The same host carrying the empty vector served as a negative control. After 24 h of incubation at 37°C, colonies were screened for fluorescence by using a fluorescence microscope as described above.

**Overlay assay to detect bioactive compounds produced by cultured isolates.** To compare the sensitivity of the intracellular screen to an overlay screen of cultured isolates, dilutions of Alaskan soil samples comparable to those used to

make the metagenomic libraries were cultured on 1/10-strength tryptic soy broth agar (i.e., 3.0 g of tryptic soy broth/liter with 1.5% agar) at 28°C for 1 week. Reporter strain JB525 was applied as an overlay to three of these plates, which each contained about 300 colonies (total of 1,000 colonies).

**Specificity of quorum-sensing inducing activity.** All assays were performed with isolated metagenomic DNA plasmids maintained in *E. coli* Transform EPI300. To determine the specificity of the active molecule with different reporter strains, overnight cultures of QS1 and QS1-C were cross-streaked with the reporter strain JB525 [*E. coli* MT102(pJBA132)], *P. putida* F117(pKR-C12), and *C. violaceum* CV026, and activity was measured after 24 h of incubation at 28°C. *E. coli* Transform EPI300 containing pSuperBAC was used as a negative control.

**Chemical characterization of culture supernatants.** Culture extracts for thin-layer chromatography (TLC) analysis were prepared from 50-ml cultures grown to stationary phase at 37°C. Bacteria were removed by centrifugation, and the supernatants were extracted twice with 50 ml of acidified ethyl acetate (0.1 ml of glacial acetic acid per liter). Extracts were evaporated to a final volume of ca. 1.5 ml and stored at -20°C. Then, 1- $\mu$ l samples were applied to C<sub>18</sub> reversed-phase TLC plates (200- $\mu$ m layer; J. T. Baker, Phillipsburg, NJ) and developed with 60:40 (vol/vol) methanol-water. Solvents were evaporated, and the plates were analyzed for the presence of acyl-HSLs with *A. tumefaciens* indicator strain KYC55(pJZ410)(pJZ384)(pJZ372). The acyl-HSL indicator was grown to stationary phase, and 5 ml of culture was added to 95 ml of AMA amended with 7.5 g of agar/liter and 50  $\mu$ g of X-Gal (5-bromo-4-chloro-3-indolyl- $\beta$ -D-galactopyranoside)/ml and then maintained at 50°C. After vortexing the culture briefly, 20 ml was applied to the TLC plate by spraying (250-ml sprayer; Kontes, Vineyard, NJ) a uniform overlay of the indicator bacteria. TLC plates were incubated at 28°C for 18 h.  $R_f$  values for compounds were compared to AHL standards, which were generous gifts from Ronald Binder (USDA ARS Pacific West Area).

**Subcloning and sequencing.** Plasmid pQS1 (from metagenomic clone QS1 that conferred fluorescence on JB525) was subjected to partial digestion with Sau3AI (Promega) and electrophoresis to size select the DNA for subcloning. Fragments greater than 2 kb were extracted from the gel and ligated into pGEM-3Zf(+) (Promega), which was linearized by using BamHI (Promega). Plasmid DNA from *E. coli* DH5 $\alpha$  transformants was prepared by the SprintPrep miniprep method (Agencourt Bioscience Corp, Beverly, MA). Purified plasmids were each sequenced by using the universal sequencing primers M13f and M13r at the University of Wisconsin DNA sequencing facility using BigDye Termina-

tor (v. 3.1; Applied Biosystems, Foster City, CA). Sequence reads were assembled by using the STADEN computer program (v. 1.4.0; <http://staden.sourceforge.net/>). Contigs and regions initially covered by only one sequence read were joined by sequencing PCR fragments until the average sequencing coverage was >7-fold and every region had at least 2-fold coverage. Putative open reading frames (ORFs) were identified by using ORF finder (<http://www.ncbi.nih.gov/gorf/gorf.html>) and were annotated by using ASAP (19). Plasmids from subclones containing metagenomic DNA were isolated, transformed into the reporter strain, plated, and screened by fluorescence microscopy to identify active subclones.

Inserts in plasmids pQS11 (from metagenomic clone QS11 that conferred fluorescence on JB525) and pINH6 (from metagenomic clone INH6 that is a putative inhibitor of fluorescence) were sequenced by primer walking from the cloning vector. ORFs were identified as described above. The pQS11 ORF of interest that had homology to isopropylmalate/homocitrate synthase was PCR amplified and cloned into pGEM-T (Promega). The ligated product was screened for GFP-inducing activity as described above.

**Phylogenetic analysis.** Phylogenetic trees of each ORF from clones QS1, QS11, and Inh6 were constructed to determine their function and closest known relative (13, 35). We obtained homologues of each ORF by performing BlastP searches of 185 finished archaeal and bacterial genomes. Those matches where the BLAST score ratio (BSR; i.e., the ratio of the BlastP score for the database match to the score of the metagenomic ORF) was more than 0.30 were retained for further analysis. This is approximately the same as 30% identity over the full length of the protein sequence. We considered any ORF without a homologue to be a hypothetical ORF. For each ORF, we used CLUSTALW to align the homologous protein sequences and constructed maximum-parsimony trees using PAUP\* (61).

The methods used to construct the LuxI-LuxR phylogenetic tree have previously been described (21, 36). Briefly, the QS1 LuxI and LuxR protein sequences were used to perform a BlastP search against the nonredundant NCBI protein database. Protein sequences from completed genomes and those from members of the *Enterobacteriaceae* were selected for further analysis. The accession numbers for the sequences used in this analysis are available online (<http://www.plantpath.wisc.edu/fac/joh/LuxIR-Accessions.htm>). The LuxI and LuxR protein sequences were concatenated (LuxIR) and aligned by using CLUSTALW (4). Any gap-containing columns in the alignment were removed from the analysis using ARB (39). Bootstrapped maximum-parsimony trees were constructed in PAUP\* version 4.0 using 1,000 replicates (61). We used MEGA version 3.0 to construct bootstrapped neighbor joining trees with 1,000 replicates and a gamma distance correction (31). Tetranucleotide frequency analysis of the insert was performed by using TETRA and its database of finished genomes (62).

**Nucleotide sequence accession numbers.** The DNA sequence of the QS1 78-kb fragment has been deposited in GenBank under accession number AY688432. The DNA sequences of the QS11 and INH6 fragments have been deposited in GenBank under accession numbers DQ008471 and DQ008472, respectively.

## RESULTS

The goal of the present study was to develop a rapid method to identify metagenomic clones that produced biologically active small molecules. We constructed large libraries of metagenomic DNA from soil and developed an intracellular screen to identify clones that induce or inhibit a bacterial quorum sensing biosensor.

**Library construction.** We constructed eight soil libraries with DNA extracted from the organic and mineral layers of soil cores from Alaska. Restriction digest analysis of 223 randomly selected clones using NotI revealed inserts ranging from 1 to 190 kb. Collectively, the libraries contain 653 Mb of DNA (Table 2).

**Biosensor.** The biosensor plasmid used in our intracellular screen is plasmid pJBA132, in which *luxR*-P<sub>luxI</sub> from *V. fischeri* is fused to *gfp*(ASV) (1). The *E. coli* host strain does not produce detectable quorum-sensing inducing compounds, and thus the GFP is expressed either when exogenous inducer is applied or a gene (or genes) directing the synthesis of an inducer is introduced into the cell. To test the specificity of the

TABLE 3. Properties of clones that induce or inhibit quorum sensing

Clone	Phenotype <sup>a</sup>	Insert size (kb)
QS1	A	78
QS1-C	A	3.5
QS4	B	24
QS5	B	42
QS6	B	29
QS8	B	11
QS9	B	30
QS10	B	10
QS11	B	3.5
QS15	B	27
QS35	B	35
QS55	B	9
INH6	C	3.7
INH11	C	9

<sup>a</sup> A, colonies fluoresced intensely after 12 h of incubation on LBA at 37°C; B, colonies fluoresced after 24 h of incubation on LBA at 37°C; C, colonies did not fluoresce after 24 h of incubation on LBA containing 3 nM *N*-(3-oxohexanoyl)-L-HSL at 37°C.

biosensor, we introduced *lasI*, *ppuI*, and *traI* from *P. aeruginosa*, *P. putida*, and *A. tumefaciens*, respectively, into the biosensor strain. The biosensor was activated by the products of each of the three genes, indicating that it detects a range of acyl-HSL compounds.

**Intracellular screen for inducers.** We inspected approximately 180,000 colonies for GFP expression by fluorescence microscopy and found three clones, QS1, QS2, and QS3, that fluoresced intensely after 12 h of incubation (Table 3). Restriction digest and pulsed-field gel electrophoresis revealed that the clones were siblings containing a 78-kb insert. The same clone was rediscovered in subsequent screens. Ten additional clones, with inserts ranging from 3.5 to 42 kb, fluoresced after 24 h of incubation (Table 3). Sequence information for pQS1 and pQS11 inserts is reported below.

Plasmids from subclones of pQS1 were transformed into the reporter strain and screened by fluorescence microscopy. QS1-C, a subclone of the plasmid in QS1, induced intense GFP in the reporter strain and carried a 3.5-kb insert whose nucleotide sequence revealed a *luxI* homologue (Fig. 2).

**Intracellular screen for inhibitors.** We used FACS to identify cells in which the presence of the metagenomic DNA results in inhibition of GFP expression induced by exogenous *N*-(3-oxohexanoyl)-L-HSL. In one screen for inhibitors of GFP expression, we included 50 nM *N*-(3-oxohexanoyl)-L-HSL and detected GFP expression on a continuum from dark to bright. In a second screen, using 80 nM *N*-(3-oxohexanoyl)-L-HSL, we detected better separation of the “dark” and “bright” phenotypes. In both screens for inhibition, approximately 10,000 cells with the “dark” phenotype were collected. To sort potential clones producing inhibitors from dead cells, the cells collected from FACS were plated onto LBA containing Cm and Tet and incubated overnight. Two clones, INH6 and INH11, had a confirmable phenotype and were selected for further study. Sequence information of pINH6 is reported below.

**Comparison of intracellular screen with overlay assay.** To determine whether the intracellular nature of the screen provided more sensitivity in detecting active clones, we tested all of the clones identified with the intracellular biosensor in a

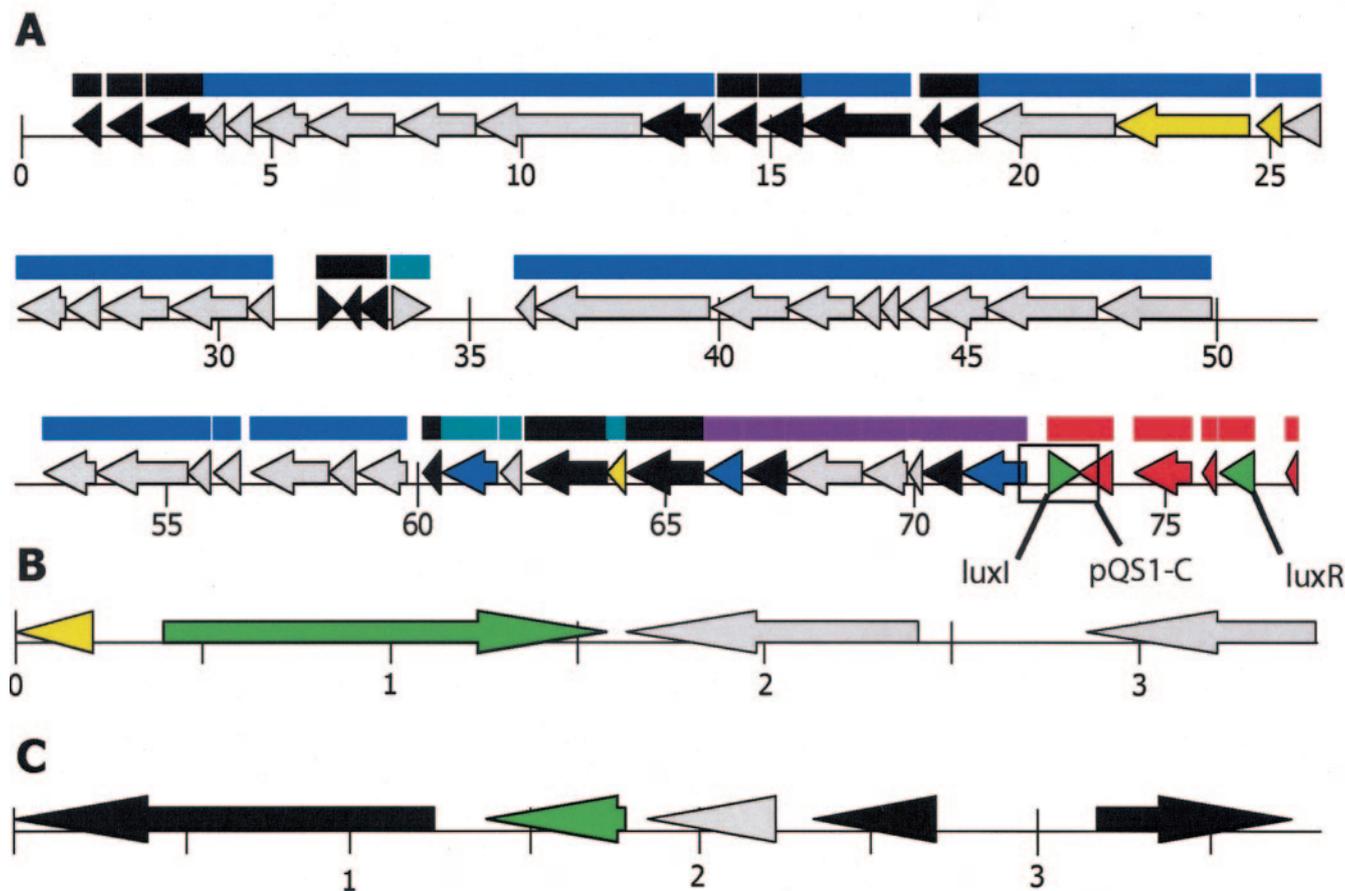


FIG. 2. Gene maps of pQS1 (A), pQS11 (B), and pINH6 (C) (distances in kb). ORFs are color coded as follows: gray, conserved hypothetical (BSR > 0.30); black, hypothetical (BSR < 0.30); yellow, ORFs with nonhypothetical BLAST matches; red, genes related to transposases; blue, genes related to acyl carrier proteins; green, *luxI* and *luxR* (A), homocitrate/isopropylmalate synthase (B), and putative cytochrome (C). The bars above the ORFs represent regions that are most similar to enterics (blue), *Chromobacterium violaceum* (purple), *Pseudomonas syringae* (red), indeterminate (turquoise), or unknown (black). The box around the *luxI* represents the region found in the subclone of pQS1 (pQS1-C) that retained the quorum-sensing activity.

typical “intercellular” or overlay screen. Among the active clones tested, only QS1 induced GFP expression when the biosensor was used as an overlay. The remaining clones (QS4 through 6, QS8 through 11, QS15, QS35, and QS55) did not activate the overlay-based reporter, indicating that the intracellular feature of our screen enhances its sensitivity and enables it to detect clones that would be missed in an overlay assay.

**Overlay assay to detect bioactive compounds produced by cultured isolates.** To compare the frequency of quorum-sensing inducing activity in metagenomic libraries with that in cultured bacteria from the same site, we tested a collection of approximately 1,000 bacterial cultures from Alaskan soil. Among these isolates, which likely contain approximately 10 times as much DNA as the metagenomic libraries, only one activated the biosensor in the overlay assay.

**Nucleotide sequence of pQS1.** We sequenced the DNA insert of pQS1 to characterize the gene responsible for the quorum-sensing phenotype and characterize its genomic environment and to reveal phylogenetic information that might indicate the affiliation of the organism from which the DNA originated (13, 35). Plasmid pQS1 contains an insert of 77,858

bp and 64 complete ORFs (Fig. 2A). A 5-kb region at the 3' end of the insert contains *luxI* and *luxR* homologues (Fig. 2A, green arrows) that are oriented toward each other and whose products reliably cluster with the *Pseudomonas* sp. group of the LuxI and LuxR homologues (Fig. 3). The LuxI homologue has 62% amino acid sequence identity to its closest match in GenBank, *AmfI* from *Pseudomonas fluorescens*. Three genes encoding putative transposases separate the *luxI* and *luxR* homologues.

The remainder of the pQS1 insert contains two large regions (27.4 and 24.5 kb, Fig. 2A, shaded in blue) in which all ORFs are most similar to ORFs from the genomes of the *Enterobacteriaceae*, except for one small region directly upstream from the *luxI* homologue that contains ORFs most similar to those of *Chromobacterium violaceum* (4.4 kb). The regions with homology to *Enterobacteriaceae* are largely syntenous with the *Y. pestis* genomes. Of the 64 ORFs, there are 35 conserved hypothetical proteins (BSR between 0.30 and 0.90), 17 are hypothetical proteins (all BSR < 0.30), 4 are putative transposases (BSR between 0.38 and 0.93), 3 are acyl-carrier proteins (BSR between 0.42 and 0.49), 1 is ferredoxin (BSR = 0.73), 1 is an ATPase subunit of an ATP-dependent protease

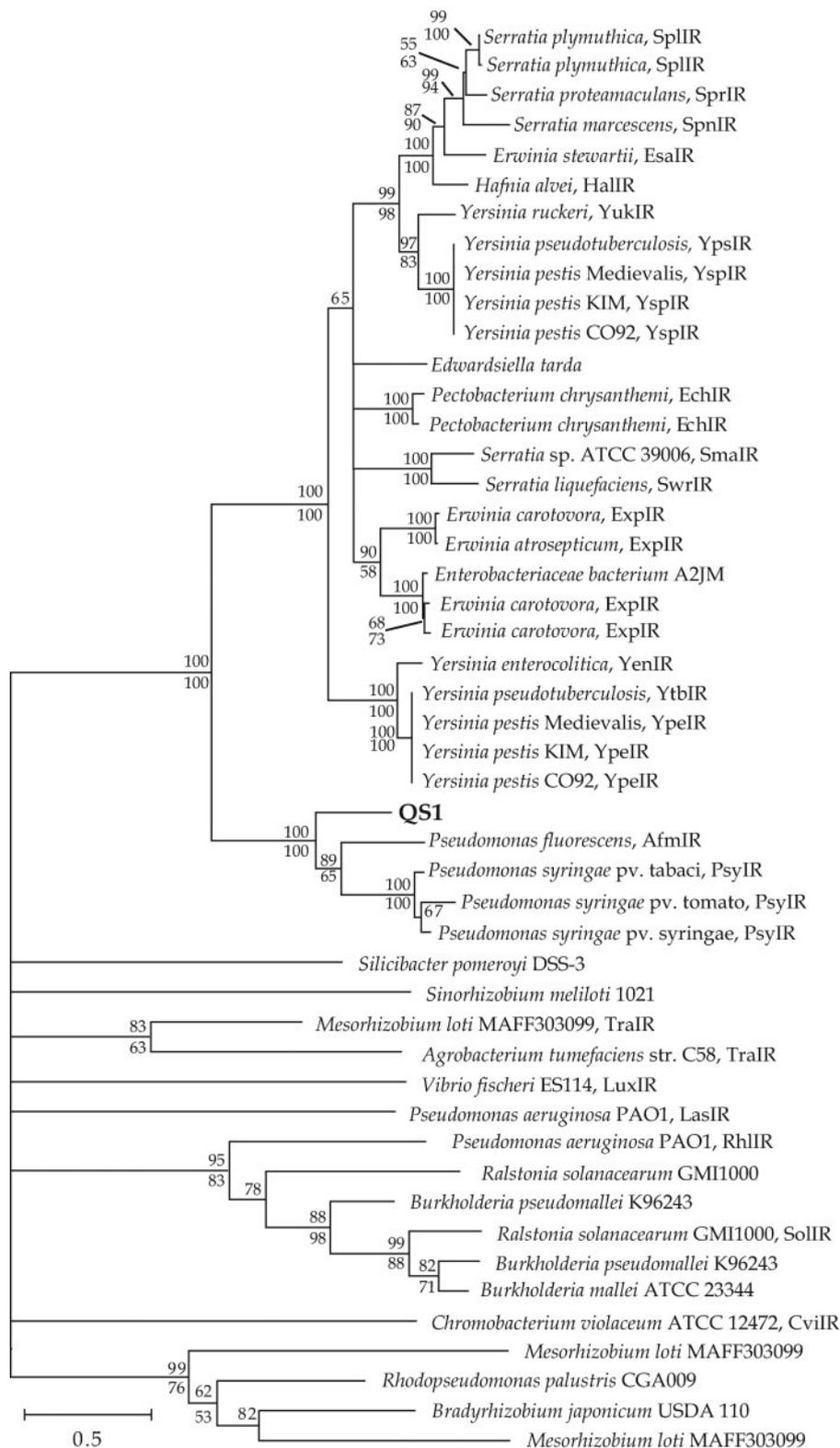


FIG. 3. Phylogenetic tree of the concatenated LuxI and LuxR protein sequence from pQS1 and its homologues. The topology of the tree was determined by using 1,000 bootstrap replicates each for neighbor joining with a gamma distance correction and using maximum parsimony. The branch lengths were taken from the neighbor-joining analysis. Numbers above and below each node represent the bootstrap values obtained by using neighbor-joining and maximum-parsimony analysis, respectively. Nodes lacking a number represent those that could not be supported by at least 50% of the bootstrap replicates. Accession numbers for each LuxI and LuxR homolog represented in the tree are available at <http://www.plantpath.wisc.edu/fac/joh/LuxIR-Accessions.htm>.

(BSR = 0.31), and 1 is a hemolysin-coregulated protein (BSR = 0.52).

Tetranucleotide frequency analysis of the ORFs in the insert contradicted the phylogenetic analysis of the ORFs. Comparison of tetranucleotide frequencies predicted that the insert was most similar to *Pseudomonas syringae* pv. tomato (correlation = 0.88). The tetranucleotide frequency correlation with the *Y. pestis* genomes was low (correlation = 0.57). When the analysis was repeated using the regions that were most similar to members of the *Enterobacteriaceae*, the *P. syringae* genome still had the highest correlation score (0.88), and correlations to the *Yersinia* sp. genomes were low (all 0.57).

The G+C content of the pQS1 insert was also analyzed. The two *Enterobacteriaceae* fragments are 55.4 and 50.8% G+C, respectively. The *C. violaceum* fragment and the region between the second *Enterobacteriaceae* fragment and the *C. violaceum* fragment are both 51% G+C. Finally, the G+C content of the *P. syringae* (i.e., the *luxI* and *luxR*) fragment is 48.3%. For comparison, the G+C content of the sequenced *P. syringae* (accession number NC\_004578) and *Y. pestis* genomes (accession numbers NC\_003143, NC\_004088, and NC\_005810) are 58 and 48%, respectively.

**Nucleotide sequence of pQS11.** Plasmid pQS11 contains a 3.5-kb insert with four ORFs. Two ORFs are hypothetical proteins, one is a fragment of a gene with homology to a transcriptional activator, and the other is similar to either 2-isopropylmalate synthase or homocitrate synthase. Protein sequence analysis of the transcriptional activator and the synthase was inconclusive because the phylogeny of the homologues for both ORFs demonstrates no consistent meaningful phylogenetic relationships. The most similar synthase homolog is from *Desulfovibrio vulgaris* (55% identity, BSR = 0.55; GenBank accession no. YP\_012192). The ORF containing the synthase gene was cloned into pGEM-T, and the resulting clone induced GFP expression in the biosensor strain.

**Nucleotide sequence of pINH6.** Plasmid pINH6 carries an insert of 3.8 kb containing five ORFs. Of the five ORFs, three are hypothetical proteins, one is 53% identical to a conserved hypothetical ORF from *Streptomyces coelicolor* (NP\_626248), and one is 32% identical to a putative cytochrome from *Lepidospira interrogans* (YP\_000668 or NP\_713687, BSR = 0.23). The mechanism of inhibition of quorum sensing by this clone is under investigation.

**Diffusion and specificity of active compounds from clones QS1 and QS1-C.** The signal molecule produced by QS1 and QS1-C diffused in agar, as indicated both by cross-streak assays, as well as when culture was spotted on a lawn of reporter strain and incubated overnight. Both clones induced high levels of expression in reporter strain JB525 and *C. violaceum* CV026, but not *P. putida* F117(pKR-C12), indicating that the clones produce compounds with similar specificity to short-chain acyl-HSLs and not to long-chain acyl-HSL molecules.

**Chemical characterization of active compounds.** The active compound(s) in QS1 and QS1-C were compared to synthetic acyl-HSLs in TLC and were detected with the overlay of the *A. tumefaciens* biosensor. The active material from QS1 and QS1-C produced teardrop-shaped patterns that were indistinguishable from the *N*-(3-oxohexanoyl)-L-HSL. Its pattern and mobility ( $R_f$  = 0.68), coupled with its biological specificity,

indicate that the active signal molecule from clone QS1 is *N*-(3-oxohexanoyl)-L-HSL.

## DISCUSSION

One of the challenges of functional metagenomics is detecting rare active clones in libraries containing many members. The potential for poor expression of genes derived from organisms distantly related to the host species harboring the library makes the development of highly sensitive assays imperative. In an attempt to address both of these issues, we developed a rapid, sensitive intracellular screen in which the sensor for activity is in the same cell as the metagenomic DNA. We designate this screen "METREX," for *metabolite-regulated expression*. We demonstrated the power of METREX in identifying metagenomic clones that produce biologically active small molecules by screening Alaskan soil microbial DNA libraries. METREX identified 11 clones that induce expression of the biosensor and two that inhibit expression. The METREX screen, conducted in host species other than *E. coli* may detect additional novel quorum-sensing genes. METREX shares certain features with the recently described screen, SIGEX (65). Whereas SIGEX is a trap for promoters that are responsive to certain metabolites, METREX detects metabolites that might activate such promoters (26, 65).

One clone identified by METREX, designated QS1, produces a biologically active small molecule that induces GFP expression. Based on its chemical properties and specificity, the signal molecule produced by QS1 appears to be an acyl-HSL, and most resembles *N*-(3-oxohexanoyl)-L-HSL. However, the gene that directs synthesis of this inducer differs from known homologues of *luxI* based on both its nucleotide sequence and the genomic organization of the flanking DNA. Based on the similarity of the 67 ORFs carried by pQS1 to other  $\gamma$ -Proteobacteria sequences and the clustering of its LuxI and LuxR homologues in a family comprised exclusively of  $\gamma$ -Proteobacteria sequences, it seems likely that this fragment originated from a member of the  $\gamma$ -Proteobacteria. Our results are consistent with studies that show acyl-HSL systems are restricted to Proteobacteria (21, 36). Further study of the 53 ORFs for which no function can be inferred based on sequence alone may provide insight into the lifestyle of the organism from which this DNA fragment originated.

Two hypotheses have been presented previously to account for the clustering of LuxI and LuxR homologues from *P. syringae* with enteric  $\gamma$ -Proteobacteria. The first suggests that bacteria similar to *P. syringae* obtained their LuxI and LuxR homologues via horizontal gene transfer from an enteric  $\gamma$ -proteobacterium (21). The second hypothesis posits two LuxI and LuxR families, one of which contains only  $\gamma$ -Proteobacteria and likely arose before the divergence of *Pseudomonas* species (36). Interestingly, the majority of the pQS1 ORFs have synteny with those of the *Y. pestis* genome (an enteric  $\gamma$ -Proteobacteria), yet the LuxIR homologues cluster with PsyIR from *P. syringae* and AmfIR from *P. fluorescens*. The tetranucleotide frequency in pQS1 is most similar to *P. syringae*. Although QS1 does not contain enough information to support either hypothesis, the presence of transposases within the LuxIR region suggests that horizontal gene transfer may have occurred.

The intracellular METREX screen detects clones that would not be detected in an overlay screen. Inhibitors and possibly inducers of the biosensor can be identified with fluorescence-activated cell sorting, which can sort 60,000 cells/min (11) and quickly identify rare active clones. Inhibitors of quorum sensing are difficult to identify in the overlay because the bright fluorescence of the majority of the colonies obscures the dark ones. Some inducers of quorum sensing would not be detectable in an overlay assay because the active compounds are not detectable outside of the producing cell. For example, ten clones that were detected by the intracellular screen did not induce GFP expression in reporter colonies when tested in an overlay assay.

An unexpected result was that the estimated frequency of quorum-sensing inducers detected in the metagenomic libraries with the intracellular screen was substantially higher than among cultured bacteria from the same soil. We found only one active colony among 1,000 colonies that were isolated from the soil, whereas we found 11 active clones in metagenomic libraries from the same soil that contained approximately 130 genome equivalents (650 Mb of DNA divided by an average genome size of 5 Mb). Although we do not know whether the difference is due to the screen (intracellular versus overlay), the source of the activity (uncultured versus cultured bacteria), or to other factors such as redundancy in culturing, the metagenomic libraries screened intracellularly appear to be richer sources of quorum sensing-inducing activity than are the cultured bacteria screened with the overlay assay.

Since the metagenomic DNA in the libraries is only a small sample of the total soil metagenome and the cultured isolates are only a small sample of the population of microbes, the frequency of quorum-sensing inducers found in either screen does not necessarily represent the proportion of quorum-sensing inducers in Alaskan soil. Both screens indicate that a small proportion of microorganisms produce detectable signals that activate the quorum-sensing biosensor. The frequency of quorum-sensing activators in soil is not known. Elasri et al. found that among *Pseudomonas* species, acyl-HSL production is common among plant-associated bacteria but uncommon among soilborne bacteria (14).

Functional metagenomics is a powerful approach for discovering new genes and gene products because it does not require prior knowledge of the gene sequence. Despite all of its limitations due to barriers of heterologous gene expression, the potential for discovery with functional metagenomics is significant, as demonstrated by the present study. With METREX, we found genes that would not have been detected with a sequence-based approach. pQS11 and pINH6, for example, do not contain genes related to any that are known to encode inducers or inhibitors of quorum sensing, and only a functional analysis could have discovered their activity. As we overcome some of the limitations of functional metagenomics, screens such as METREX will become more fruitful and may help realize the promise of metagenomics for discovery of small molecules.

#### ACKNOWLEDGMENTS

We thank Michael Givskov for the reporter strain JB525, Ronald Binder for the acyl-HSL standards, Leo Eberl for F117(pKR-C12), and Steve Winans for the *A. tumefaciens* acyl-HSL indicator strain. We also thank Zakee Sabree for designing Fig. 1 and Nicole T. Perna for facilitating our use of ASAP.

P.D.S. was partially supported by a USDA soil biology postdoctoral fellowship. This study was supported by grants from the National Science Foundation Microbial Observatory program (MCB-0132085), The Howard Hughes Medical Institute Professors Program, the Biotechnology Research and Development Corporation, and Hatch Project 4534 from the College of Agricultural and Life Sciences of the University of Wisconsin–Madison.

#### REFERENCES

- Andersen, J. B., A. Heydorn, M. Hentzer, L. Eberl, O. Geisenberger, B. B. Christensen, S. Molin, and M. Givskov. 2001. *gfp*-based *N*-acyl homoserine-lactone sensor systems for detection of bacterial communication. *Appl. Environ. Microbiol.* **67**:575–585.
- Bainton, N. J., P. Stead, S. R. Chhabra, B. W. Bycroft, G. P. Salmond, G. S. Stewart, and P. Williams. 1992. *N*-(3-oxohexanoyl)-L-homoserine lactone regulates carbapenem antibiotic production in *Erwinia carotovora*. *Biochem. J.* **288**:997–1004.
- Beck von Bodman, S., G. T. Hayman, and S. K. Farrand. 1992. Opine catabolism and conjugal transfer of the nopaline Ti plasmid pTiC58 are coordinately regulated by a single repressor. *Proc. Natl. Acad. Sci. USA* **89**:643–647.
- Chenna, R., H. Sugawara, T. Koike, R. Lopez, T. J. Gibson, D. G. Higgins, and J. D. Thompson. 2003. Multiple sequence alignment with the CLUSTAL series of programs. *Nucleic Acids Res.* **31**:3497–3500.
- Cho, K., C. Fuqua, and S. C. Winans. 1997. Transcriptional regulation and locations of *Agrobacterium tumefaciens* genes required for complete catabolism of octopine. *J. Bacteriol.* **179**:1–8.
- Courtois, S., C. M. Cappellano, M. Ball, F. X. Francou, P. Normand, G. Helynyck, A. Martinez, S. J. Kolvek, J. Hopke, M. S. Osburne, P. R. August, R. Nalin, M. Guerinou, P. Jeannin, P. Simonet, and J. L. Pernodet. 2003. Recombinant environmental libraries provide access to microbial diversity for drug discovery from natural products. *Appl. Environ. Microbiol.* **69**:49–55.
- Curtis, T. P., W. T. Sloan, and J. W. Scannell. 2002. Estimating prokaryotic diversity and its limits. *Proc. Natl. Acad. Sci. USA* **99**:10494–10499.
- Daniel, R. 2004. The soil metagenome: a rich resource for the discovery of novel natural products. *Curr. Opin. Biotechnol.* **15**:199–204.
- Degrassi, G., C. Aguilar, M. Bosco, S. Zahariev, S. Pongor, and V. Venturi. 2002. Plant growth-promoting *Pseudomonas putida* WCS358 produces and secretes four cyclic dipeptides: cross-talk with quorum sensing bacterial sensors. *Curr. Microbiol.* **45**:250–254.
- Dong, Y. H., J. L. Xu, X. Z. Li, and L. H. Zhang. 2000. AiiA, an enzyme that inactivates the acylhomoserine lactone quorum-sensing signal and attenuates the virulence of *Erwinia carotovora*. *Proc. Natl. Acad. Sci. USA* **97**:3526–3531.
- Dunn, A. K., and J. Handelsman. 1999. A vector for promoter trapping in *Bacillus cereus*. *Gene* **226**:297–305.
- Eberhard, A. A., L. Burlingame, C. Eberhard, G. L. Kenyon, K. H. Neilson, and N. J. Oppenheimer. 1981. Structural identification of autoinducer of *Photobacterium fischeri* luciferase. *Biochemistry* **20**:2444–2449.
- Eisen, J. A. 1998. Phylogenomics: improving functional predictions for uncharacterized genes by evolutionary analysis. *Genome Res.* **8**:163–167.
- Elasri, M., S. Delorme, P. Lemanceau, G. Stewart, B. Laue, E. Glickmann, P. M. Oger, and Y. Dessaux. 2001. Acyl-homoserine lactone production is more common among plant-associated *Pseudomonas* spp. than among soilborne *Pseudomonas* spp. *Appl. Environ. Microbiol.* **67**:1198–1209.
- Fuqua, W. C., S. C. Winans, and E. P. Greenberg. 1994. Quorum sensing in bacteria: the LuxR-LuxI family of cell density-responsive transcriptional regulators. *J. Bacteriol.* **176**:269–275.
- Gao, M., M. Teplitski, J. B. Robinson, and W. D. Bauer. 2003. Production of substances by *Medicago truncatula* that affect bacterial quorum sensing. *Mol. Plant-Microbe Interact.* **16**:827–834.
- Gillespie, D. E., S. F. Brady, A. D. Bettermann, N. P. Cianciotto, M. R. Liles, M. R. Rondon, J. Clardy, R. M. Goodman, and J. Handelsman. 2002. Isolation of antibiotics turbinocin A and B from a metagenomic library of soil microbial DNA. *Appl. Environ. Microbiol.* **68**:4301–4306.
- Givskov, M., R. de Nys, M. Manefield, L. Gram, R. Maximilien, L. Eberl, S. Molin, P. D. Steinberg, and S. Kjelleberg. 1996. Eukaryotic interference with homoserine lactone-mediated prokaryotic signaling. *J. Bacteriol.* **178**:6618–6622.
- Glasner, J. D., P. Liss, G. Plunkett III, A. Darling, T. Prasad, M. Rusch, A. Byrnes, M. Gilson, B. Biehl, F. R. Blattner, and N. T. Perna. 2003. ASAP, a systematic annotation package for community analysis of genomes. *Nucleic Acids Res.* **31**:147–151.
- Goh, E. B., G. Yim, W. Tsui, J. McClure, M. G. Surette, and J. Davies. 2002. Transcriptional modulation of bacterial gene expression by subinhibitory concentrations of antibiotics. *Proc. Natl. Acad. Sci. USA* **99**:17025–17030.
- Gray, K. M., and J. R. Garey. 2001. The evolution of bacterial LuxI and LuxR quorum sensing regulators. *Microbiology* **147**:2379–2387.
- Handelsman, J., M. R. Rondon, S. F. Brady, J. Clardy, and R. M. Goodman.

1998. Molecular biological access to the chemistry of unknown soil microbes: a new frontier for natural products. *Chem. Biol.* **5**:R245–R249.
23. Handelsman, J., M. R. Liles, D. A. Mann, C. S. Riesenfeld, and R. M. Goodman. 2002. Cloning the metagenome: culture-independent access to the diversity and functions of the uncultivated microbial world, p. 241–255. *In* B. W. A. N. Dorrell (ed.), *Functional microbial genomics*. Academic Press, Inc., New York, N.Y.
  24. Handelsman, J. 2004. Metagenomics: application of genomics to uncultured microorganisms. *Microbiol. Mol. Biol. Rev.* **68**:669–685.
  25. Handelsman, J. 2004. Soils: the metagenomics approach, p. 109–120. *In* A. T. Bull (ed.), *Microbial diversity and bioprospecting*. ASM Press, Washington, D.C.
  26. Handelsman, J. 2005. Sorting out metagenomes. *Nat. Biotechnol.* **23**:38–39.
  27. Hastings, J. W., and K. H. Neelson. 1977. Bacterial bioluminescence. *Annu. Rev. Microbiol.* **31**:549–595.
  28. Holden, M. T., S. Ram Chhabra, R. de Nys, P. Stead, N. J. Bainton, P. J. Hill, M. Manefield, N. Kumar, M. Labatte, D. England, S. Rice, M. Givskov, G. P. Salmond, G. S. Stewart, B. W. Bycroft, S. Kjelleberg, and P. Williams. 1999. Quorum-sensing cross talk: isolation and chemical characterization of cyclic dipeptides from *Pseudomonas aeruginosa* and other gram-negative bacteria. *Mol. Microbiol.* **33**:1254–1266.
  29. Hwang, I., P. L. Li, L. Zhang, K. R. Piper, D. M. Cook, M. E. Tate, and S. K. Farrand. 1994. TraI, a LuxI homologue, is responsible for production of conjugation factor, the Ti plasmid *N*-acylhomoserine lactone autoinducer. *Proc. Natl. Acad. Sci. USA* **91**:4639–4643.
  30. Jones, S., B. Yu, N. J. Bainton, M. Birdsall, B. W. Bycroft, S. R. Chhabra, A. J. Cox, P. Golby, P. J. Reeves, S. Stephens, et al. 1993. The *lux* autoinducer regulates the production of exoenzyme virulence determinants in *Erwinia carotovora* and *Pseudomonas aeruginosa*. *EMBO J.* **12**:2477–2482.
  31. Kumar, S., K. Tamura, and M. Nei. 2004. MEGA3: integrated software for molecular evolutionary genetics analysis and sequence alignment. *Brief Bioinform.* **5**:150–163.
  32. Latifi, A., M. K. Winson, M. Foglino, B. W. Bycroft, G. S. Stewart, A. Lazdunski, and P. Williams. 1995. Multiple homologues of LuxR and LuxI control expression of virulence determinants and secondary metabolites through quorum sensing in *Pseudomonas aeruginosa* PAO1. *Mol. Microbiol.* **17**:333–343.
  33. Leadbetter, J. R., and E. P. Greenberg. 2000. Metabolism of acyl-homoserine lactone quorum-sensing signals by *Variovorax paradoxus*. *J. Bacteriol.* **182**:6921–6926.
  34. Leps, W. T., W. J. Brill, and E. T. Bingham. 1980. Effect of alfalfa ploidy on nitrogen fixation. *Crop Sci.* **20**:427–430.
  35. Lerat, E., V. Daubin, and N. A. Moran. 2003. From gene trees to organismal phylogeny in prokaryotes: the case of the gamma-Proteobacteria. *PLoS Biol.* **1**:E19.
  36. Lerat, E., and N. A. Moran. 2004. The evolutionary history of quorum-sensing systems in bacteria. *Mol. Biol. Evol.* **21**:903–913.
  37. Liles, M. R., L. L. Williamson, J. Handelsman, and R. M. Goodman. 2004. Isolation of high-molecular-weight genomic DNA from soil bacteria for genomic library construction, p. 839–850. *In* G. A. Kowalchuk, F. J. de Bruijn, I. M. Head, A. D. L. Akkermans, and J. D. van Elsas (ed.), *Molecular microbial ecology manual*, 2nd ed. Kluwer Academic Publishers, Dordrecht, The Netherlands.
  38. Lorenz, P., K. Liebeton, F. Niehaus, and J. Eck. 2002. Screening for novel enzymes for biocatalytic processes: accessing the metagenome as a resource of novel functional sequence space. *Curr. Opin. Biotechnol.* **13**:572–577.
  39. Ludwig, W., O. Strunk, R. Westram, L. Richter, H. Meier, Yadhukumar, A. Buchner, T. Lai, S. Steppi, G. Jobb, W. Forster, I. Brettske, S. Gerber, A. W. Ginhart, O. Gross, S. Grumann, S. Hermann, R. Jost, A. Konig, T. Liss, R. Lussmann, M. May, B. Nonhoff, B. Reichel, R. Strehlow, A. Stamatakis, N. Stuckmann, A. Vilbig, M. Lenke, T. Ludwig, A. Bode, and K. H. Schleifer. 2004. ARB: a software environment for sequence data. *Nucleic Acids Res.* **32**:1363–1371.
  40. Manefield, M., T. B. Rasmussen, M. Henzter, J. B. Andersen, P. Steinberg, S. Kjelleberg, and M. Givskov. 2002. Halogenated furanones inhibit quorum sensing through accelerated LuxR turnover. *Microbiology* **148**:1119–1127.
  41. McClean, K. H., M. K. Winson, L. Fish, A. Taylor, S. R. Chhabra, M. Camara, M. Daykin, J. H. Lamb, S. Swift, B. W. Bycroft, G. S. Stewart, and P. Williams. 1997. Quorum sensing and *Chromobacterium violaceum*: exploitation of violacein production and inhibition for the detection of *N*-acylhomoserine lactones. *Microbiology* **143**:3703–3711.
  42. McFarland, J. W., R. W. Ruess, K. Kielland, and A. P. Doyle. 2002. Cycling dynamics of NH<sub>4</sub><sup>+</sup> and amino acid nitrogen in soils of a deciduous boreal forest ecosystem. *Ecosystems* **5**:775–788.
  43. McGowan, S., M. Sebahia, S. Jones, B. Yu, N. Bainton, P. F. Chan, B. Bycroft, G. S. Stewart, P. Williams, and G. P. Salmond. 1995. Carbapenem antibiotic production in *Erwinia carotovora* is regulated by CarR, a homologue of the LuxR transcriptional activator. *Microbiology* **141**(Pt. 3):541–550.
  44. Neelson, K. H., T. Platt, and J. W. Hastings. 1970. Cellular control of the synthesis and activity of the bacterial luminescent system. *J. Bacteriol.* **104**:313–322.
  45. Omura, S. 1992. Thom Award Lecture: trends in the search for bioactive microbial metabolites. *J. Ind. Microbiol.* **10**:135–156.
  46. Paradkar, A., A. Trefzer, R. Chakraborty, and D. Stassi. 2003. *Streptomyces* genetics: a genomic perspective. *Crit. Rev. Biotechnol.* **23**:1–27.
  47. Passador, L., J. M. Cook, M. J. Gambello, L. Rust, and B. H. Iglewski. 1993. Expression of *Pseudomonas aeruginosa* virulence genes requires cell-to-cell communication. *Science* **260**:1127–1130.
  48. Pettit, R. K. 2004. Soil DNA libraries for anticancer drug discovery. *Cancer Chemother. Pharmacol.* **54**:1–6.
  49. Pirhonen, M., D. Flego, R. Heikinheimo, and E. T. Palva. 1993. A small diffusible signal molecule is responsible for the global control of virulence and exoenzyme production in the plant pathogen *Erwinia carotovora*. *EMBO J.* **12**:2467–2476.
  50. Riesenfeld, C. S., P. D. Schloss, and J. Handelsman. 2004. Metagenomics: genomic analysis of microbial communities. *Annu. Rev. Genet.* **38**:525–552.
  51. Rivkina, E. M., E. I. Friedmann, C. P. McKay, and D. A. Gilichinsky. 2000. Metabolic activity of permafrost bacteria below the freezing point. *Appl. Environ. Microbiol.* **66**:3230–3233.
  52. Rondon, M. R., P. R. August, A. D. Bettermann, S. F. Brady, T. H. Grossman, M. R. Liles, K. A. Loiacono, B. A. Lynch, I. A. MacNeil, C. Minor, C. L. Tiong, M. Gilman, M. S. Osburne, J. Clardy, J. Handelsman, and R. M. Goodman. 2000. Cloning the soil metagenome: a strategy for accessing the genetic and functional diversity of uncultured microorganisms. *Appl. Environ. Microbiol.* **66**:2541–2547.
  53. Sambrook, J., and D. W. Russell. 2001. *Molecular cloning: a laboratory manual*, 3rd ed. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y.
  54. Schloss, P. D., and J. Handelsman. 2003. Biotechnological prospects from metagenomics. *Curr. Opin. Biotechnol.* **14**:303–310.
  55. Segawa, T., K. Miyamoto, K. Ushida, K. Agata, N. Okada, and S. Kohshima. 2005. Seasonal change in bacterial flora and biomass in mountain snow from the Tateyama Mountains, Japan, analyzed by 16S rRNA gene sequencing and real-time PCR. *Appl. Environ. Microbiol.* **71**:123–130.
  56. Skidmore, M. L., J. M. Foght, and M. J. Sharp. 2000. Microbial life beneath a high arctic glacier. *Appl. Environ. Microbiol.* **66**:3214–3220.
  57. Steidle, A., K. Sigl, R. Schuegger, A. Ihring, M. Schmid, S. Gantner, M. Stoffels, K. Riedel, M. Givskov, A. Hartmann, C. Langebartels, and L. Eberl. 2001. Visualization of *N*-acylhomoserine lactone-mediated cell-cell communication between bacteria colonizing the tomato rhizosphere. *Appl. Environ. Microbiol.* **67**:5761–5770.
  58. Steidle, A., M. Allesen-Holm, K. Riedel, G. Berg, M. Givskov, S. Molin, and L. Eberl. 2002. Identification and characterization of an *N*-acylhomoserine lactone-dependent quorum-sensing system in *Pseudomonas putida* strain IsoF. *Appl. Environ. Microbiol.* **68**:6371–6382.
  59. Stover, C. K., X. Q. Pham, A. L. Erwin, S. D. Mizoguchi, P. Warren, M. J. Hickey, F. S. Brinkman, W. O. Hufnagle, D. J. Kowalik, M. Lagrou, R. L. Garber, L. Goltry, E. Tolentino, S. Westbrook-Wadman, Y. Yuan, L. L. Brody, S. N. Coulter, K. R. Folger, A. Kas, K. Larbig, R. Lim, K. Smith, D. Spencer, G. K. Wong, Z. Wu, I. T. Paulsen, J. Reizer, M. H. Saier, R. E. Hancock, S. Lory, and M. V. Olson. 2000. Complete genome sequence of *Pseudomonas aeruginosa* PAO1, an opportunistic pathogen. *Nature* **406**:959–964.
  60. Streit, W. R., R. Daniel, and K. E. Jaeger. 2004. Prospecting for biocatalysts and drugs in the genomes of noncultured microorganisms. *Curr. Opin. Biotechnol.* **15**:285–290.
  61. Swofford, D. L. 2003. PAUP\*: phylogenetic analysis using parsimony (\* and other methods), version 4. Sinauer Associates, Sunderland, Mass.
  62. Teeling, H., J. Waldmann, T. Lombardot, M. Bauer, and F. O. Glockner. 2004. TETRA: a web-service and a stand-alone program for the analysis and comparison of tetranucleotide usage patterns in DNA sequences. *BMC Bioinformatics* **5**:163.
  63. Torsvik, V., J. Goksoyr, and F. L. Daae. 1990. High diversity in DNA of soil bacteria. *Appl. Environ. Microbiol.* **56**:782–787.
  64. Torsvik, V. 1995. Cell extraction method, p. 1.3.1/1-1.3.1/15. *In* A. D. L. Akkermans, J. D. Van Elsas, and F. J. de Bruijn (ed.), *Molecular microbial ecology manual*. Kluwer Academic Publishers, Dordrecht, The Netherlands.
  65. Uchiyama, T., T. Abe, T. Ikemura, and K. Watanabe. 2005. Substrate-induced gene-expression screening of environmental metagenome libraries for isolation of catabolic genes. *Nat. Biotechnol.* **23**:88–93.
  66. von Bodman, S. B., D. R. Majerczak, and D. L. Coplin. 1998. A negative regulator mediates quorum-sensing control of exopolysaccharide production in *Pantoea stewartii* subsp. *stewartii*. *Proc. Natl. Acad. Sci. USA* **95**:7687–7692.
  67. Wood, D. W., and L. S. Pierson III. 1996. The *phzI* gene of *Pseudomonas aureofaciens* 30-84 is responsible for the production of a diffusible signal required for phenazine antibiotic production. *Gene* **168**:49–53.
  68. Zhou, J., M. A. Bruns, and J. M. Tiedje. 1996. DNA recovery from soils of diverse composition. *Appl. Environ. Microbiol.* **62**:316–322.
  69. Zhou, J., M. E. Davey, J. B. Figueras, E. Rivkina, D. Gilichinsky, and J. M. Tiedje. 1997. Phylogenetic diversity of a bacterial community determined from Siberian tundra soil DNA. *Microbiology* **143**:3913–3919.
  70. Zhu, J., Y. Chai, Z. Zhong, S. Li, and S. C. Winans. 2003. *Agrobacterium* bioassay strain for ultrasensitive detection of *N*-acylhomoserine lactone-type quorum-sensing molecules: detection of autoinducers in *Mesorhizobium huakuii*. *Appl. Environ. Microbiol.* **69**:6949–6953.